

Review

Melatonin As a Protective Agent Against Environmental Stresses: A Review into Its Molecular Regulation in Plants

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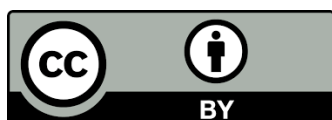
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Abstract

Understanding the impact of melatonin (N-acetyl-5-methoxytryptamine) on plant gene expression is crucial for unlocking its full potential as a tool for crop improvement and stress tolerance. Melatonin has emerged to have several influences on the transcriptional activity of numerous genes, helping to orchestrate plant responses to environmental cues. Furthermore, it has been shown that melatonin signaling pathways control downstream gene expression to ensure proper plant growth and development. Therefore, clearing out the complex interaction between melatonin and plant gene expression has enormous potential to further our knowledge of plant biology and develop novel farming techniques.



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In this review, we have gathered the recent studies that elucidate the role of applied melatonin in regulating stress-responsive genes under various abiotic stresses.

Keywords

Melatonin; abiotic stress; gene expression and transcription factors

1. Introduction

Melatonin, commonly known as the "sleep hormone," is an endogenous hormone that regulates the sleep-wake cycle in animals and humans and is produced by the pineal gland [1]. However, it has been found that melatonin is also vital in plants, specifically in gene expression that regulates various plant metabolic, growth, and developmental processes [2-4]. Melatonin can be a signaling molecule regulating plant growth and stress responses [5]. For example, melatonin can upregulate gene expression involved in cell division, ethylene, and isoflavones biosynthesis in *Glycine max* [6]. Furthermore, melatonin has been discovered to promote the production of stress-responsive genes, which helps plants cope with diverse environmental challenges such as drought, salinity, and temperature extremes [7-10].

On the other hand, melatonin has been found to interact with other plant hormones, i.e., auxin [11], ABA [12], polyamines [7], and cytokinin [13]. In addition, multiple lines of evidence suggest that melatonin can affect the expression of genes involved in ROS detoxification and antioxidant defense systems, leading to plant survival under several stressful conditions [3, 14-16]. Furthermore, melatonin is vital in controlling plant gene expression by acting as a transcriptional regulator, affecting various physiological processes [17]. For example, melatonin has been shown to enhance the expression of genes involved in stress tolerance, photosynthesis, and root development while suppressing the expression of genes involved in senescence and cell death [18-20].

It is also involved in epigenetic regulation, which refers to heritable changes in gene expression without any changes in the DNA sequence itself [21]. In this respect, Ahmad *et al.* [21] found that melatonin can influence DNA methylation and histone modifications, which are essential in the epigenetic mechanisms regulating gene expression. Through this modulation of epigenetic marks, melatonin can alter the accessibility of genes to transcription factors, resulting in modifications to gene expression patterns [16]. This review endeavors to understand the molecular mechanisms by which melatonin helps plants cope with various environmental challenges, emphasizing its potential as a protective agent (Figure 1).

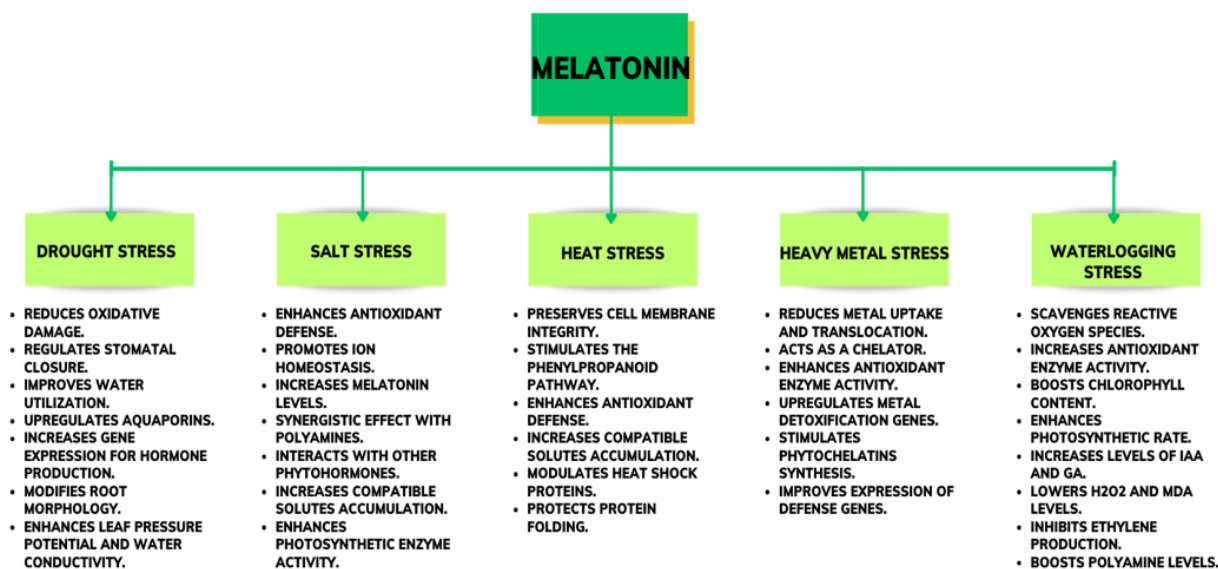


Figure 1 Melatonin acts as a protective agent against various environmental stresses.

2. Melatonin and Delaying of Leaf Senescence

Leaf senescence, or the final stage of leaf development, substantially impacts plant productivity and survival. Melatonin has been shown to slow this process, extending the functional lifespan of leaves. Melatonin has been known as a promoter of the synthesis of photosynthetic pigments in plants. Studies have shown that melatonin can enhance chlorophyll and carotenoid levels in leaves, promoting photosynthetic activity and improving plant growth and development [12, 22, 23]. Melatonin can reduce the produced free radicals by scavenging various cascade reactions and inducing several antioxidative defense systems [10, 12, 24]. It has been found that melatonin can prevent photoinhibition in chloroplast by accelerating the non-photochemical quenching, which protects the structure of photosynthetic proteins and lipids and promotes the xanthophyll cycle [25]. Research suggests that melatonin inhibits the breakdown of chlorophyll by reducing the expression of genes associated with chlorophyll degradation, including pheophorbide an oxygenase (PAO), pheophytinase (PPH), red chlorophyll catabolite reductase (RCCR), 7-hydroxymethyl chlorophyll a reductase (HCAR), Non-Yellowing Coloring 1 like (NOL), and Non-Yellowing Coloring 1 (NYC1). Additionally, melatonin restricts the activity of enzymes involved in chlorophyll catabolism, such as chlorophyllase (CLH) and PPH [26-28]. Wu *et al.* [28] found that treating broccoli (*Brassica oleracea* L. var. Italica, cv. Youxiu) with melatonin caused a down-regulation in the expression of genes responsible for breaking down chlorophyll. These genes included NYC1, NOL, CLH, PPH, PAO, RCCR, and Stay-Green 1 (SGR1). Furthermore, melatonin has been found to be essential in controlling the transcriptional reprogramming of senescing leaves, i.e., NACs, WRKYs, and DREBs [20]. In another study, Martinez *et al.* [29] found that the application of melatonin to apple plants resulted in a delay in leaf senescence. This delay was achieved by inhibiting the expression of specific genes involved in chlorophyll degradation, including notably senescence-associated gene 12 (SAG12) and auxin-resistant 3 (AXR3)/indole-3-acetic acid inducible 17 (IAA17). Moving on from the topic of leaf senescence, it is critical to investigate how melatonin's protective properties apply to various environmental stressors, beginning with drought stress.

3. Melatonin and Alleviating of Drought Stress

Drought stress has a severe impact on plant growth and yield. Drought stress is a major environmental factor that limits crop productivity and threatens global food security [30]. It has been found that applying melatonin externally can reduce oxidative damage caused by drought, regulate stomata closure, and improve water utilization efficiency in plants [2, 22, 31]. Under water deficit conditions, melatonin regulates the expression of various genes responsible for producing plant hormones such as ABA, IAA, GAs, CKs, ethylene jasmonic acid (JA), and brassinosteroids. This regulation occurs by increasing the expression of hormone receptors, associated signaling elements, and biosynthesis enzymes [32]. Moreover, melatonin has been found to upregulate specific aquaporins, membrane proteins responsible for water transport, facilitating water uptake and its distribution throughout the plant tissues [32]. In this respect, it has been found that exogenous melatonin enhanced the leaf pressure potential (Lpr) and the water conductivity of the plant (K_{plant}) in maize plants by upregulating PIP aquaporin genes, which are responsible for water transport in the presence of drought stress [33]. It was also reported that the rhizospheric application of melatonin increases the mRNA expression level of TIP aquaporins and prevents water loss in barley [34]. According to Xia *et al.* [35], melatonin can also increase the expression levels of the MAPKs *Asmap1* and *Aspk11*, as well as the *WRKY1*, *DREB2*, and *MYB* transcription factor genes in oat seedlings under drought stress. These findings suggest that activating the MAPK cascade may be necessary for melatonin to induce the antioxidant response and regulate the expression of genes relevant to antioxidants. Besides, melatonin has been reported to modify root morphology and promote root elongation, enabling plants to explore deeper soil layers with higher water availability [36]. On the other hand, melatonin has been shown to regulate stomatal closure during drought stress, allowing plants to strike a balance between water conservation and efficient CO₂ fixation [37]. This modulation occurs through melatonin-mediated regulation of abscisic acid (ABA), an essential hormone involved in stomatal movement, suggesting a potential interplay between melatonin and other signaling molecules to fine-tune stomatal responses and optimize plant water-use efficiency [38]. After discussing drought stress, we will look at how melatonin might assist in combating salt stress, another common environmental challenge.

4. Melatonin and Alleviating of Salt Stress

Salt stress impairs plant health by creating ionic imbalance and osmotic stress. Exogenous melatonin has been found to mediate salt tolerance in plants by several interactions with MAP kinase signaling pathways. In this regard, melatonin has been found to upregulate significantly the expression of antioxidant enzyme genes, nicotinamide adenine dinucleotide phosphate (NADPH) oxidase genes, mitogen-activated protein kinase (MAPK) genes (*MAPK3*, *MAPK4*, *MAPK6*) and salt overly sensitive (SOS) genes (*SOS1*, *SOS2*, *SOS3*) in salt-stressed cucumber plants [39]. Furthermore, melatonin can promote ion homeostasis, specifically the Na/K ratio under salt stress. This response is associated with the upregulation of several genes, such as *NHX*, *SOS*, and *AKT* [40]. Li *et al.* [41] found that exogenous melatonin significantly increased the expression of *MdNHX1* and *MdAKT1* genes in the salt-stressed seedlings of *Malus hupehensis*. This observation was in line with elevating the levels of potassium (K⁺) and the ratio of K⁺ to sodium (Na⁺) in plant tissues. On the other hand, exogenous melatonin can induce an elevation in endogenous melatonin

levels through activation of the phyto-melatonin receptor CAND2/PMTR1 [41]. Melatonin has been found to synergize with polyamines and reduce their catabolism in snap bean seedlings under salinity stress, leading to multiple protective effects against salt stress [7]. Melatonin also interacts with the other phytohormones by affecting their biosynthetic-related genes. For example, exogenous melatonin has been found to down-regulate *NCED1* and *CsNCED2*, which serve as ABA synthesis-related genes. At the same time, ABA catabolism-related genes and GA biosynthesis-related genes (*GA20ox* and *GA3ox*) were upregulated [40]. One key mechanism through which melatonin modulates salt stress is by enhancing antioxidant defense systems [42]. In this context, Zhang *et al.* [43] found that exogenous melatonin increased the expression of genes associated with the removal of reactive oxygen species (ROS), including *CsCu-ZnSOD*, *CsFe-ZnSOD*, *CsPOD*, and *CsCAT* in salt-stressed cucumber seedlings. Moreover, melatonin promotes the accumulation of compatible solutes, such as proline and free amino acids and total soluble sugars, which protect plants from osmotic stress caused by high salt concentration [7, 38, 44]. Additionally, previous studies have shown that melatonin enhances the activity of critical enzymes that are involved in photosynthesis, such as Rubisco and ATP synthase, thus ensuring efficient energy production in the presence of salt stress by enhancing the efficiency of the electron transport chain, which increases ATP production and thereby facilitates plant resilience under salt stress [45, 46]. Temperature extremes, in addition to salt stress, are a critical stress factor that affects plant health. Let's look at how melatonin helps with both low and high-temperature stress.

5. Melatonin and Alleviating of Heat Stress: Low and High Temperatures

Extreme temperatures, both high and low, could significantly impact plant metabolism and growth. Low temperatures (chilling stress) significantly impact plant cell membrane integrity, critical for preserving cellular functioning [47]. The physical characteristics of the membrane's lipids lose some of their fluidity and may go through a phase change, increasing its vulnerability to cold injury [48-50]. In a previous study on tomato fruits during cold storage, exogenous melatonin (100 μ M) has been found to decrease the chilling injury by supplying enough intracellular ATP and enhancing the activities of H⁺-ATPase, Ca-ATPase, cytochrome C oxidase (CCO), and succinate dehydrogenase (SDH) [51]. At the same time, these responses were associated with preserving the integrity of the cellular membranes by achieving a higher ratio of unsaturated to saturated fatty acids by increasing the expression of the *FAD3* and *FAD7* genes and decreasing *PLD* and *LOX* genes [51]. Moreover, applied melatonin has been found to stimulate the phenylpropanoid pathway by promoting the activities of phenylalanine ammonia-lyase (PAL), 4-coumarate-coenzyme A ligase (4CL), cinnamate-4-hydroxylase (4CH) and peroxidase (POD), and accompanied by higher contents of total phenols and lignin, which might be contributed to improving the temperature tolerance in plum fruit during storage [52]. The antioxidant property of melatonin can also prevent cellular damage and maintain cell membranes' structural and functional integrity under low-temperature conditions [53-56].

On the other hand, high temperatures have detrimental effects on different physiological processes, decreasing plant growth and even leading to cell death [57]. It can disrupt plant water balance, leading to dehydration and wilting [58]. Melatonin has been found to enhance the accumulation of compatible solutes, such as proline and soluble sugars, which act as osmoprotectants under heat stress [3, 14, 59]. Also, applied melatonin can increase plant

thermotolerance by modulating the expression of heat shock proteins (HSPs) [60]. In this context, melatonin significantly stimulated the expression of the carotenoid biosynthesis gene in the presence of 10 *de novo* HSPs in kiwifruit [61]. Furthermore, melatonin can protect plant cells from heat-induced damage by affecting protein folding and preventing its denaturation [62].

Additionally, melatonin has been demonstrated to increase the antioxidative capacity of *Camellia sinensis* L under heat stress by increasing the transcript levels of catechins biosynthesis genes, including *CsCHS*, *CsCH1*, *CsF3H*, *CsDFR*, *CsANS*, *CsLAR*, and *CsANR* [63]. In addition to temperature, contamination by heavy metals in soils is another significant stressor. The following section explores how melatonin helps plants deal with heavy metal stress.

6. Melatonin and Alleviating of Heavy Metal Stress

Heavy metal stress occurs when harmful metals accumulate in plant tissues, inducing oxidative damage and diminished physiological functioning. Heavy metals are toxic elements that can accumulate in plants and pose serious health risks to humans and ecosystems [64]. Melatonin has been shown to play a crucial role in regulating heavy metal accumulation in plants [65]. Research suggests that melatonin can mitigate the harmful effects of heavy metals by reducing metal uptake and translocation within the plant [66]. This hormone acts as a chelator, binding to heavy metals and forming less toxic complexes and more easily excreted [67].

Additionally, melatonin can enhance the activity of antioxidant enzymes, which help combat the oxidative stress induced by heavy metal toxicity [68]. By regulating metal accumulation, melatonin contributes to plants' overall health and resilience in metal-contaminated environments. Furthermore, melatonin possesses a unique ability to induce metal detoxification mechanisms in plants. Studies have found that melatonin can upregulate the expression of genes involved in metal detoxification, including metallothioneins and transporters that facilitate metal sequestration in vacuoles [69]. This upregulation enhances the plant's ability to cope with heavy metal stress and reduces the toxic effects caused by metal accumulation in vital plant tissues.

Moreover, melatonin can stimulate the synthesis of phytochelatin, small proteins that bind to heavy metals, and enhance their sequestration and detoxification within plants [70]. In a previous study on copper toxicity in tomato seedlings, applied melatonin improved the expression of several defense genes (CAT, APX, GR, and MDHAR) and melatonin biosynthesis-related genes, i.e., *TDC*, *SNAT*, and *COMT* [71]. Under nickel toxicity, Jahan *et al.* [26] found that tomato seedlings treated with melatonin effectively inhibited the generation of hydrogen peroxide (H₂O₂) and superoxide radicals. It also raised the expression of *RBOH* and restored cellular integrity by reducing malondialdehyde levels and electrolyte leakage. This effect was achieved by activating antioxidant enzymes and regulating the AsA-GSH pools. The study found that nickel-induced oxidative stress was successfully reduced by upregulating the expression of many defense genes (*SOD*, *CAT*, *APX*, *GR*, *GST*, *MDHAR*, *DHAR*) and genes associated with melatonin production (*TDC*, *T5S*, *SNAT*, *ASMT*). Yet another stressor to consider is waterlogging, which may also harm plant life. The next part will address how melatonin assists in alleviating waterlogging stress.

7. Melatonin and Alleviating Waterlogging

Plants under wet conditions experience hypoxia or low oxygen levels, which set off a series of physiological and biochemical reactions [72]. Melatonin uses a variety of regulatory systems to

assist plants in lessening the negative consequences of waterlogging. Plants experience oxidative stress due to the overproduction of reactive oxygen species (ROS) brought on by waterlogging. Melatonin, a potent antioxidant, is essential for scavenging ROS and decreasing oxidative damage in a variety of plant species, including wheat [73], alfalfa [74], kiwifruit [75], and maize [76]. Previous research showed that the MDA content of maize seedlings decreased when melatonin was used under waterlogging stress [76]. Several studies have indicated that melatonin increases the activity of antioxidant enzymes like SOD, POD, and CAT [22, 77]. In recent work on cotton grown under waterlogging, melatonin also boosted leaf chlorophyll content, photosynthetic rate, IAA and gibberellic acid (GA) levels, as well as SOD, POD, and CAT activity [72]. Melatonin, on the other hand, lowered the concentrations of H₂O₂, MDA, and ABA, as well as the activities of alcohol dehydrogenase (ADH) and pyruvate decarboxylase (PDC), when compared to waterlogging without using melatonin [72].

Additionally, melatonin increased the expression of the melatonin biosynthesis genes *GhSNAT1* and *GhCOMT* and the gibberellin biosynthesis gene *GhGA20ox1*, compared to non-waterlogged cotton. Melatonin, on the other hand, reduced the expression of the ABA synthesis gene *GhNCED2*, the H₂O₂ generating gene *GhRBOHC*, and the glycolysis and fermentation gene *GhADH2* [72]. In alfalfa, melatonin inhibited ethylene production by downregulating ethylene biosynthesis-related genes and preventing waterlogging-induced growth decrease, chlorosis, and early senescence in plants. Also, melatonin boosts polyamine levels by increasing polyamine metabolism enzymes' activity and gene expression [74]. These reactions allow plants to adapt to and endure extended durations of water saturation.

8. Conclusion and Future Prospective

In conclusion, melatonin is a crucial hormonal regulator of gene expression in plants, playing a significant role in plant development, growth, and stress response. Its ability to interact with transcription factors and modulate epigenetic mechanisms gives plants a dynamic and flexible way to adapt to changing environmental conditions. Besides, the molecular mechanisms behind melatonin's plant activities are still unknown. Despite enormous advances, more extensive investigations are needed to explain the mechanisms involved and identify melatonin's direct targets in plant cells. Further research into the specific genes and pathways regulated by melatonin in plants will uncover more insights into its role in plant biology. Additional research is required to fully understand the intricate molecular mechanisms underlying this relationship. Still, the findings offer promising avenues for remediation strategies in metal-contaminated soils and environments. By harnessing melatonin's potential, we may develop innovative approaches to mitigate heavy metal toxicity in plants and promote ecological sustainability. Another point of contention is whether melatonin occurs naturally in plants. Some researchers wonder whether the quantities of melatonin found in plants can produce the documented protective effects or if the high doses utilized in experiments artificially enhance the molecule's influence. Still, the possible ecological and environmental consequences of broad melatonin use in agriculture are in doubt. The long-term implications on plant health, soil microbiology, and overall ecosystem balance should be further investigated. By addressing these unaddressed issues and improving our understanding of melatonin's mechanisms, we can better take advantage of its benefits to boost plant resilience and productivity in agriculture.

Author Contributions

B. O. S. Al-Falahi, I. Lamdjad conceived the idea; M. Al-Nujaifi and N. A. Alheety visualisation, A. Qayyum resources B. O. S. Al-Falahi, funding B. O. S. Al-Falahi and I. Lamdjad, revision; B. O. S. Al-Falahi, I. Lamdjad wrote the draft of manuscript, A. Qayyum supervision; all authors contributed to writing the manuscript.

Competing Interests

The authors have declared that no competing interests exist.

References

1. Arendt J, Aulinas A. Physiology of the pineal gland and melatonin. Endotext. South Dartmouth, MA: MDText.com, Inc.; 2000.
2. Colombage R, Singh MB, Bhalla PL. Melatonin and abiotic stress tolerance in crop plants. *Int J Mol Sci.* 2023; 24: 7447.
3. Jahan MS, Guo S, Sun J, Shu S, Wang Y, Abou El-Yazied A, et al. Melatonin-mediated photosynthetic performance of tomato seedlings under high-temperature stress. *Plant Physiol Biochem.* 2021; 167: 309-320.
4. Raja V, Qadir SU, Kumar N, Alsahli AA, Rinklebe J, Ahmad P. Melatonin and strigolactone mitigate chromium toxicity through modulation of ascorbate-glutathione pathway and gene expression in tomato. *Plant Physiol Biochem.* 2023; 201: 107872.
5. Gao Y, Chen H, Chen D, Hao G. Genetic and evolutionary dissection of melatonin response signaling facilitates the regulation of plant growth and stress responses. *J Pineal Res.* 2023; 74: e12850.
6. Kumar G, Saad KR, Arya M, Puthusseri B, Mahadevappa P, Shetty NP, et al. The synergistic role of serotonin and melatonin during temperature stress in promoting cell division, ethylene and isoflavones biosynthesis in *Glycine max*. *Curr Plant Biol.* 2021; 26: 100206.
7. El-Beltagi HS, El-Yazied AA, El-Gawad HG, Kandeel M, Shalaby TA, Mansour AT, et al. Synergistic impact of melatonin and putrescine interaction in mitigating salinity stress in snap bean seedlings: Reduction of oxidative damage and inhibition of polyamine catabolism. *Horticulturae.* 2023; 9: 285.
8. Jan R, Asif S, Asaf S, Du XX, Park JR, Nari K, et al. Melatonin alleviates arsenic (As) toxicity in rice plants via modulating antioxidant defense system and secondary metabolites and reducing oxidative stress. *Environ Pollut.* 2023; 318: 120868.
9. Jiang D, Lu B, Liu L, Duan W, Meng Y, Li J, et al. Exogenous melatonin improves the salt tolerance of cotton by removing active oxygen and protecting photosynthetic organs. *BMC Plant Biol.* 2021; 21: 331.
10. Sun C, Meng S, Wang B, Zhao S, Liu Y, Qi M, et al. Exogenous melatonin enhances tomato heat resistance by regulating photosynthetic electron flux and maintaining ROS homeostasis. *Plant Physiol Biochem.* 2023; 196: 197-209.
11. Zia SF, Berkowitz O, Bedon F, Whelan J, Franks AE, Plummer KM. Direct comparison of *Arabidopsis* gene expression reveals different responses to melatonin versus auxin. *BMC Plant Biol.* 2019; 19: 567.

12. El-Yazied AA, Ibrahim MF, Ibrahim MA, Nasef IN, Al-Qahtani SM, Al-Harbi NA, et al. Melatonin mitigates drought induced oxidative stress in potato plants through modulation of osmolytes, sugar metabolism, ABA homeostasis and antioxidant enzymes. *Plants*. 2022; 11: 1151.
13. Wang Y, Li J, Yang L, Chan Z. Melatonin antagonizes cytokinin responses to stimulate root growth in *Arabidopsis*. *J Plant Growth Regul*. 2023; 42: 1833-1845.
14. Jahan MS, Shu S, Wang Y, Hasan MM, El-Yazied AA, Alabdallah NM, et al. Melatonin pretreatment confers heat tolerance and repression of heat-induced senescence in tomato through the modulation of ABA-and GA-mediated pathways. *Front Plant Sci*. 2021; 12: 650955.
15. Jahan MS, Zhao CJ, Shi LB, Liang XR, Jabborova D, Nasar J, et al. Physiological mechanism of melatonin attenuating to osmotic stress tolerance in soybean seedlings. *Front Plant Sci*. 2023; 14: 1193666.
16. Kaya C, Ugurlar F. Melatonin and stress tolerance in horticultural crops: Insights into gene regulation, epigenetic modifications, and hormonal interplay. *Sci Hortic*. 2023; 322: 112432.
17. Wei Y, Liu G, Chang Y, Lin D, Reiter RJ, He C, et al. Melatonin biosynthesis enzymes recruit WRKY transcription factors to regulate melatonin accumulation and transcriptional activity on W-box in cassava. *J Pineal Res*. 2018; 65: e12487.
18. Kobylńska A, Reiter RJ, Posmyk MM. Melatonin protects cultured tobacco cells against lead-induced cell death via inhibition of cytochrome c translocation. *Front Plant Sci*. 2017; 8: 1560.
19. Zhao D, Wang H, Chen S, Yu D, Reiter RJ. Phytomelatonin: An emerging regulator of plant biotic stress resistance. *Trends Plant Sci*. 2021; 26: 70-82.
20. Zhao YQ, Zhang ZW, Chen YE, Ding CB, Yuan S, Reiter RJ, et al. Melatonin: A potential agent in delaying leaf senescence. *CRC Crit Rev Plant Sci*. 2021; 40: 1-22.
21. Ahmad N, Naeem M, Ali H, Alabbosh KF, Hussain H, Khan I, et al. From challenges to solutions: The impact of melatonin on abiotic stress synergies in horticultural plants via redox regulation and epigenetic signaling. *Sci Hortic*. 2023; 321: 112369.
22. Ibrahim MF, Elbar OH, Farag R, Hikal M, El-Kelish A, El-Yazied AA, et al. Melatonin counteracts drought induced oxidative damage and stimulates growth, productivity and fruit quality properties of tomato plants. *Plants*. 2020; 9: 1276.
23. Nie M, Ning N, Chen J, Zhang Y, Li S, Zheng L, et al. Melatonin enhances salt tolerance in sorghum by modulating photosynthetic performance, osmoregulation, antioxidant defense, and ion homeostasis. *Open Life Sci*. 2023; 18: 20220734.
24. Arnao MB, Hernández-Ruiz J. Melatonin: Plant growth regulator and/or biostimulator during stress? *Trends Plant Sci*. 2014; 19: 789-797.
25. Ding F, Wang M, Liu B, Zhang S. Exogenous melatonin mitigates photoinhibition by accelerating non-photochemical quenching in tomato seedlings exposed to moderate light during chilling. *Front Plant Sci*. 2017; 8: 244.
26. Jahan MS, Guo S, Baloch AR, Sun J, Shu S, Wang Y, et al. Melatonin alleviates nickel phytotoxicity by improving photosynthesis, secondary metabolism and oxidative stress tolerance in tomato seedlings. *Ecotoxicol Environ Saf*. 2020; 197: 110593.
27. Szafrńska K, Reiter RJ, Posmyk MM. Melatonin improves the photosynthetic apparatus in pea leaves stressed by paraquat via chlorophyll breakdown regulation and its accelerated de novo synthesis. *Front Plant Sci*. 2017; 8: 878.

28. Wu C, Cao S, Xie K, Chi Z, Wang J, Wang H, et al. Melatonin delays yellowing of broccoli during storage by regulating chlorophyll catabolism and maintaining chloroplast ultrastructure. *Postharvest Biol Technol.* 2021; 172: 111378.
29. Martinez V, Nieves-Cordones M, Lopez-Delacalle M, Rodenas R, Mestre TC, Garcia-Sanchez F, et al. Tolerance to stress combination in tomato plants: New insights in the protective role of melatonin. *Molecules.* 2018; 23: 535.
30. Kang Y, Khan S, Ma X. Climate change impacts on crop yield, crop water productivity and food security-A review. *Prog Nat Sci.* 2009; 19: 1665-1674.
31. Huang B, Chen YE, Zhao YQ, Ding CB, Liao JQ, Hu C, et al. Exogenous melatonin alleviates oxidative damages and protects photosystem II in maize seedlings under drought stress. *Front Plant Sci.* 2019; 10: 677.
32. Tiwari RK, Lal MK, Kumar R, Chourasia KN, Naga KC, Kumar D, et al. Mechanistic insights on melatonin-mediated drought stress mitigation in plants. *Physiol Plant.* 2021; 172: 1212-1226.
33. Qiao Y, Ren J, Yin L, Liu Y, Deng X, Liu P, et al. Exogenous melatonin alleviates PEG-induced short-term water deficiency in maize by increasing hydraulic conductance. *BMC Plant Biol.* 2020; 20: 218.
34. Kurowska MM, Wiecha K, Gajek K, Szarejko I. Drought stress and re-watering affect the abundance of TIP aquaporin transcripts in barley. *PLoS One.* 2019; 14: e0226423.
35. Xia H, Ni Z, Hu R, Lin L, Deng H, Wang J, et al. Melatonin alleviates drought stress by a non-enzymatic and enzymatic antioxidative system in kiwifruit seedlings. *Int J Mol Sci.* 2020; 21: 852.
36. Altaf MA, Shahid R, Ren MX, Naz S, Altaf MM, Khan LU, et al. Melatonin improves drought stress tolerance of tomato by modulating plant growth, root architecture, photosynthesis, and antioxidant defense system. *Antioxidants.* 2022; 11: 309.
37. Jensen NB, Ottosen CO, Zhou R. Exogenous melatonin alters stomatal regulation in tomato seedlings subjected to combined heat and drought stress through mechanisms distinct from ABA signaling. *Plants.* 2023; 12: 1156.
38. Ali M, Pan Y, Liu H, Cheng Z. Melatonin interaction with abscisic acid in the regulation of abiotic stress in Solanaceae family plants. *Front Plant Sci.* 2023; 14: 1271137.
39. Zhang T, Shi Z, Zhang X, Zheng S, Wang J, Mo J. Alleviating effects of exogenous melatonin on salt stress in cucumber. *Sci Hortic.* 2020; 262: 109070.
40. Zhan H, Nie X, Zhang T, Li S, Wang X, Du X, et al. Melatonin: A small molecule but important for salt stress tolerance in plants. *Int J Mol Sci.* 2019; 20: 709.
41. Li C, Wang P, Wei Z, Liang D, Liu C, Yin L, et al. The mitigation effects of exogenous melatonin on salinity-induced stress in *malus hupehensis*. *J Pineal Res.* 2012; 53: 298-306.
42. Li J, Liu J, Zhu T, Zhao C, Li L, Chen M. The role of melatonin in salt stress responses. *Int J Mol Sci.* 2019; 20: 1735.
43. Zhang HJ, Zhang NA, Yang RC, Wang L, Sun QQ, Li DB, et al. Melatonin promotes seed germination under high salinity by regulating antioxidant systems, ABA and GA 4 interaction in cucumber (*Cucumis sativus* L.). *J Pineal Res.* 2014; 57: 269-279.
44. Ali M, Kamran M, Abbasi GH, Saleem MH, Ahmad S, Parveen A, et al. Melatonin-induced salinity tolerance by ameliorating osmotic and oxidative stress in the seedlings of two tomato (*Solanum lycopersicum* L.) cultivars. *J Plant Growth Regul.* 2021; 40: 2236-2248.

45. Qu C, Liu C, Ze Y, Gong X, Hong M, Wang L, et al. Inhibition of nitrogen and photosynthetic carbon assimilation of maize seedlings by exposure to a combination of salt stress and potassium-deficient stress. *Biol Trace Elem Res.* 2011; 144: 1159-1174.
46. Teng Z, Zheng W, Jiang S, Hong SB, Zhu Z, Zang Y. Role of melatonin in promoting plant growth by regulating carbon assimilation and ATP accumulation. *Plant Sci.* 2022; 319: 111276.
47. Zahra N, Shaukat K, Hafeez MB, Raza A, Hussain S, Chaudhary MT, et al. Physiological and molecular responses to high, chilling, and freezing temperature in plant growth and production: Consequences and mitigation possibilities. In: *Harsh environment and plant resilience: Molecular and functional aspects.* Cham: Springer; 2021. pp. 235-290.
48. Fontanier C, Moss JQ, Gopinath L, Goad C, Su K, Wu Y. Lipid composition of three bermudagrasses in response to chilling stress. *J Am Soc Hortic Sci.* 2020; 145: 95-103.
49. Mazur R, Gieczewska K, Kowalewska Ł, Kuta A, Proboszcz M, Gruszecki WI, et al. Specific composition of lipid phases allows retaining an optimal thylakoid membrane fluidity in plant response to low-temperature treatment. *Front Plant Sci.* 2020; 11: 723.
50. Venzhik Y, Deryabin A, Moshkov I. Adaptive strategy of plant cells during chilling: Aspect of ultrastructural reorganization. *Plant Sci.* 2023; 332: 111722.
51. Jannatizadeh A, Aghdam MS, Luo Z, Razavi F. Impact of exogenous melatonin application on chilling injury in tomato fruits during cold storage. *Food Bioproc Tech.* 2019; 12: 741-750.
52. Yan R, Xu Q, Dong J, Kebbeh M, Shen S, Huan C, et al. Effects of exogenous melatonin on ripening and decay incidence in plums (*Prunus salicina* L. cv. Taoxingli) during storage at room temperature. *Sci Hortic.* 2022; 292: 110655.
53. Jiao J, Jin M, Liu H, Suo J, Yin X, Zhu Q, et al. Application of melatonin in kiwifruit (*Actinidia chinensis*) alleviated chilling injury during cold storage. *Sci Hortic.* 2022; 296: 110876.
54. Song L, Tan Z, Zhang W, Li Q, Jiang Z, Shen S, et al. Exogenous melatonin improves the chilling tolerance and preharvest fruit shelf life in eggplant by affecting ROS-and senescence-related processes. *Hortic Plant J.* 2023; 9: 523-540.
55. Wang M, Zhang S, Ding F. Melatonin mitigates chilling-induced oxidative stress and photosynthesis inhibition in tomato plants. *Antioxidants.* 2020; 9: 218.
56. Zhang X, Feng Y, Jing T, Liu X, Ai X, Bi H. Melatonin promotes the chilling tolerance of cucumber seedlings by regulating antioxidant system and relieving photoinhibition. *Front Plant Sci.* 2021; 12: 789617.
57. Distéfano AM, Martin MV, Córdoba JP, Bellido AM, D'Ippólito S, Colman SL, et al. Heat stress induces ferroptosis-like cell death in plants. *J Cell Biol.* 2017; 216: 463-476.
58. Hassan MU, Chattha MU, Khan I, Chattha MB, Barbanti L, Aamer M, et al. Heat stress in cultivated plants: Nature, impact, mechanisms, and mitigation strategies-A review. *Plant Biosyst Int J Deal Aspects Plant Biosyst.* 2021; 155: 211-234.
59. Li ZG, Xu Y, Bai LK, Zhang SY, Wang Y. Melatonin enhances thermotolerance of maize seedlings (*Zea mays* L.) by modulating antioxidant defense, methylglyoxal detoxification, and osmoregulation systems. *Protoplasma.* 2019; 256: 471-490.
60. Raza A, Charagh S, García-Caparrós P, Rahman MA, Ogwugwa VH, Saeed F, et al. Melatonin-mediated temperature stress tolerance in plants. *GM Crops Food.* 2022; 13: 196-217.
61. Xia H, Zhou Y, Deng H, Lin L, Deng Q, Wang J, et al. Melatonin improves heat tolerance in *Actinidia deliciosa* via carotenoid biosynthesis and heat shock proteins expression. *Physiol Plant.* 2021; 172: 1582-1593.

62. Xu W, Cai SY, Zhang Y, Wang Y, Ahammed GJ, Xia XJ, et al. Melatonin enhances thermotolerance by promoting cellular protein protection in tomato plants. *J Pineal Res.* 2016; 61: 457-469.
63. Li X, Li MH, Deng WW, Ahammed GJ, Wei JP, Yan P, et al. Exogenous melatonin improves tea quality under moderate high temperatures by increasing epigallocatechin-3-gallate and theanine biosynthesis in *camellia sinensis* L. *J Plant Physiol.* 2020; 253: 153273.
64. Tumanyan AF, Seliverstova AP, Zaitseva NA. Effect of heavy metals on ecosystems. *Chem Technol Fuels Oils.* 2020; 56: 390-394.
65. Hoque MN, Tahjib-Ul-Arif M, Hannan A, Sultana N, Akhter S, Hasanuzzaman M, et al. Melatonin modulates plant tolerance to heavy metal stress: Morphological responses to molecular mechanisms. *Int J Mol Sci.* 2021; 22: 11445.
66. Ou C, Cheng W, Wang Z, Yao X, Yang S. Exogenous melatonin enhances Cd stress tolerance in *Platycladus orientalis* seedlings by improving mineral nutrient uptake and oxidative stress. *Ecotoxicol Environ Saf.* 2023; 252: 114619.
67. Galano A, Medina ME, Tan DX, Reiter RJ. Melatonin and its metabolites as copper chelating agents and their role in inhibiting oxidative stress: A physicochemical analysis. *J Pineal Res.* 2015; 58: 107-116.
68. Yang H, Fang R, Luo L, Yang W, Huang Q, Yang C, et al. Potential roles of melatonin in mitigating the heavy metals toxicity in horticultural plants. *Sci Hortic.* 2023; 321: 112269.
69. Tang M, Xu L, Wang Y, Dong J, Zhang X, Wang K, et al. Melatonin-induced DNA demethylation of metal transporters and antioxidant genes alleviates lead stress in radish plants. *Hortic Res.* 2021; 8: 124. doi: 10.1038/s41438-021-00561-8.
70. Xing Q, Hasan MK, Li Z, Yang T, Jin W, Qi Z, et al. Melatonin-induced plant adaptation to cadmium stress involves enhanced phytochelatin synthesis and nutrient homeostasis in *solanum lycopersicum* L. *J Hazard Mater.* 2023; 456: 131670.
71. Zhang T, Wang Y, Ma X, Ouyang Z, Deng L, Shen S, et al. Melatonin alleviates copper toxicity via improving ROS metabolism and antioxidant defense response in tomato seedlings. *Antioxidants.* 2022; 11: 758.
72. Zhang Y, Liang T, Dong H. Melatonin enhances waterlogging tolerance of field-grown cotton through quiescence adaptation and compensatory growth strategies. *Field Crops Res.* 2024; 306: 109217.
73. Ma S, Gai P, Geng B, Wang Y, Ullah N, Zhang W, et al. Exogenous melatonin improves waterlogging tolerance in wheat through promoting antioxidant enzymatic activity and carbon assimilation. *Agronomy.* 2022; 12: 2876.
74. Zhang Q, Liu X, Zhang Z, Liu N, Li D, Hu L. Melatonin improved waterlogging tolerance in alfalfa (*Medicago sativa*) by reprogramming polyamine and ethylene metabolism. *Front Plant Sci.* 2019; 10: 44.
75. Huo L, Wang H, Wang Q, Gao Y, Xu K, Sun X. Exogenous treatment with melatonin enhances waterlogging tolerance of kiwifruit plants. *Front Plant Sci.* 2022; 13: 1081787.
76. Ahmad S, Wang GY, Muhammad I, Zeeshan M, Zhou XB. Melatonin and KNO₃ application improves growth, physiological and biochemical characteristics of maize seedlings under waterlogging stress conditions. *Biology.* 2022; 11: 99.

77. Ali MA, Nasser MA, Abdelhamid AN, Ali IA, Saady HS, Hassan KM. Melatonin as a key factor for regulating and relieving abiotic stresses in harmony with phytohormones in horticultural plants-a review. *J Soil Sci Plant Nutr.* 2023; 24: 54-73.